

ANTIMICROBIAL CONTACT LENSES AND METHODS FOR THEIR
PRODUCTION
RELATED INVENTIONS

This patent application claims priority from a provisional patent
5 application, U.S. Ser. No. 60/257,030, that was filed on December 21, 2000.

FIELD OF THE INVENTION

This invention relates to contact lenses having antimicrobial properties as
well as methods of their production, use, and storage.

BACKGROUND OF THE INVENTION

Contact lenses have been used commercially to improve vision since the
1950s. The first contact lenses were made of hard materials. Although these
lenses are currently used, they are not suitable for all patients due to their poor
15 initial comfort and their relatively low permeability to oxygen. Later
developments in the field gave rise to soft contact lenses, based upon hydrogels,
which are extremely popular today. Many users find soft lenses are more
comfortable, and increased comfort levels allow soft contact lens users to wear
their lenses for far longer hours than users of hard contact lenses.

20 Despite this advantage, the extended use of the lenses can encourage
the buildup of bacteria or other microbes, particularly, *Pseudomonas aeruginosa*,
on the surfaces of soft contact lenses. The build-up of bacteria or other
microbes is not unique to soft contact lens wearers and may occur during the
use of hard contact lenses as well.

25 Therefore, there is a need to produce contact lenses that inhibit the
growth of bacteria or other microbes and/or the adhesion of bacterial or other
microbes on the surface of contact lenses. Further there is a need to produce
contact lenses which do not promote the adhesion and/or growth of bacteria or
other microbes on the surface of the contact lenses. Also there is a need to
30 produce contact lenses that inhibit adverse responses related to the growth of
bacteria or other microbes.

Others have recognized the need to produce soft contact lenses that inhibit the growth of bacteria. In US Patent No. 5,213,801, the production of an antibacterial contact lens is disclosed, where an antibacterial metal ceramic material within a soft contact lens is incorporated into a contact lens. This procedure contains a number of steps and may not be suitable for producing all types of lenses in a production environment. The steps include making a silver ceramic material that is fine enough to be used in a contact lens and then forming the lens with the powdered ceramic. However, lenses containing these types of materials often lack the clarity required by contact lens users.

Although these methods and lenses are known, other contact lenses that inhibit the growth and/or adhesion of bacteria or other microbes and are of sufficient optical clarity, as well as methods of making those lenses are still needed. It is this need, which this invention seeks to meet.

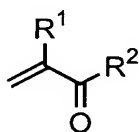
BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 Lenses N & G Movement and Silver Concentration

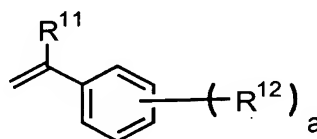
Figure 2 Lens Q Movement and Silver Concentration

DETAILED DESCRIPTION OF THE INVENTION

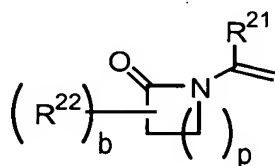
This invention includes an antimicrobial lens comprising, consisting essentially of, or consisting of, silver and a polymer comprising a monomer of Formula I, II, III or IV



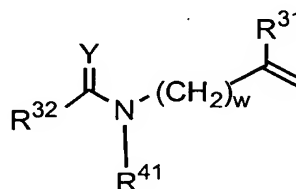
I



II



III



IV

wherein

R^1 is hydrogen or C_{1-6} alkyl;

R^2 is $-OR^3$, $-NH-R^3$, $-S-(CH_2)_d-R^3$, or $-(CH_2)_d-R^3$, wherein

d is 0-8;

R^3 is substituted C_{1-6} alkyl

where the alkyl substituents are selected from one or more members of the group consisting of carboxylic acid, sulfonic acid, phosphonic acid, amine, amidine, acetamide, nitrile, thiol, C_{1-6} alkyldisulfide, C_{1-6} alkylsulfide, phenyldisulfide, urea, C_{1-6} alkylurea, phenylurea, thiourea, C_{1-6} alkylthiourea, phenylthiourea, substituted C_{1-6} alkyldisulfide, substituted phenyldisulfide, substituted C_{1-6} alkylurea, substituted phenylurea, substituted C_{1-6} alkylthiourea, and substituted phenylthiourea

wherein the C_{1-6} alkyldisulfide, phenyldisulfide, C_{1-6} alkylurea, C_{1-6} alkylthiourea, phenylurea, and phenylthiourea substituents are selected from the group consisting of C_{1-6} alkyl, halo C_{1-6} alkyl, halogen, hydroxyl, carboxylic acid, sulfonic acid, phosphonic acid, amine, amidine, acetamide, and nitrile;

$-(CR^4R^5)_q-(CHR^6)_m-SO_3H$

wherein R^4 , R^5 , and R^6 are independently selected from the group consisting of hydrogen, halogen, hydroxyl, and C_{1-6} alkyl,

q is 1-6, and

m is 0-6;

$-(CH_2)_n-S-S-(CH_2)_x-NH-C(O)CR^7CH_2$,

wherein R^7 is hydrogen or C_{1-6} alkyl,

n is 1-6, and

x is 1-6;



wherein R^8 , R^9 , and R^{10} are independently selected from the group consisting of hydrogen, halogen, hydroxyl, and C_{1-6} alkyl,

t is 1-6, and

u is 0-6;

phenyl;

benzyl;

pyridinyl;

pyrimidinyl;

pyrazinyl;

benzimidazolyl;

benzothiazolyl;

benzotriazolyl;

naphthaloyl;

quinolinyl;

indolyl;

thiadiazolyl;

triazolyl;

4-methylpiperidin-1-yl;

4-methylpiperazin-1-yl;

substituted phenyl;

substituted benzyl;

substituted pyridinyl;

substituted pyrimidinyl;

substituted pyrazinyl;

substituted benzimidazolyl;

substituted benzothiazolyl;

substituted benzotriazolyl;

substituted naphthaloyl;

substituted quinolinyl;

substituted indolyl;
substituted thiadiazolyl;
substituted triazolyl;
substituted 4-methylpiperidin-1-yl; or
substituted 4-methylpiperazin-1-yl,

wherein the substituents are selected from one or more members of the group consisting of C₁₋₆alkyl, haloC₁₋₆alkyl, halogen, sulfonic acid, phosphonic acid, hydroxyl, carboxylic acid, amine, amidine, N-(2-aminopyrimidine)sulfonyl, N-(aminopyridine)sulfonyl, N-(aminopyrazine)sulfonyl, N-(2-aminopyrimidine)carbonyl, N-(aminopyridine)carbonyl, N-(aminopyrazine)carbonyl, N-(2-aminopyrimidine)phosphonyl, N-(2-aminopyridine)phosphonyl, N-(aminopyrazine)phosphonyl, N-(aminobenzimidazolyl)sulfonyl, N-(aminobenzothiazolyl)sulfonyl, N-(aminobenzotriazolyl)sulfonyl, N-(aminoindolyl)sulfonyl, N-(aminothiazolyl)sulfonyl, N-(aminotriazolyl)sulfonyl, N-(amino-4-methylpiperidinyl)sulfonyl, N-(amino-4-methylpiperazinyl)sulfonyl, N-(aminobenzimidazolyl)carbonyl, N-(aminobenzothiazolyl)carbonyl, N-(aminobenzotriazolyl)carbonyl, N-(aminoindolyl)carbonyl, N-(aminothiazolyl)carbonyl, N-(aminotriazolyl)carbonyl, N-(amino-4-methylpiperidinyl)carbonyl, N-(amino-4-methylpiperazinyl)carbonyl, N-(2-aminobenzimidazolyl)phosphonyl,

N-(2-aminobenzothiazolyl)phosphonyl,
 N-(2-aminobenzotriazolyl)phosphonyl,
 N-(2-aminoindolyl)phosphonyl,
 N-(2-aminothiazolyl)phosphonyl,
 N-(2-aminotriazolyl)phosphonyl,
 N-(amino-4-methylpiperidiny) phosphonyl,
 N-(amino-4-methylpiperaziny) phosphonyl, acetamide,
 nitrile, thiol, C₁₋₆alkyldisulfide, C₁₋₆alkylsulfide, phenyl
 disulfide, urea, C₁₋₆alkylurea, phenylurea, thiourea,
 C₁₋₆alkylthiourea, phenylthiourea, substituted
 C₁₋₆alkyldisulfide, substituted phenyldisulfide, substituted
 C₁₋₆alkylurea, substituted C₁₋₆alkylthiourea, substituted
 phenylurea, and substituted phenylthiourea
 wherein the C₁₋₆alkyldisulfide, phenyldisulfide,
 C₁₋₆alkylurea, C₁₋₆alkylthiourea, phenylurea, and
 phenylthiourea substituents are selected from the
 group consisting of C₁₋₆alkyl, haloC₁₋₆alkyl, halogen,
 hydroxyl, carboxylic acid, sulfonic acid, phosphonic
 acid, amine, amidine, acetamide, and nitrile;

a is 1-5;

R¹¹ is hydrogen or C₁₋₆alkyl;

R¹² is hydroxyl, sulfonic acid, phosphonic acid, carboxylic acid,
 acetamide, thioC₁₋₆alkylcarbonyl, C₁₋₆alkyldisulfide, C₁₋₆alkylsulfide,
 phenyl disulfide, urea, C₁₋₆alkylurea, phenylurea, thiourea,
 C₁₋₆alkylthiourea, phenylthiourea, -OR¹³, -NH-R¹³, -S-(CH₂)_d-R¹³,
 -(CH₂)_d-R¹³, -C(O)NH--(CH₂)_d-R¹³, -C(O)-(CH₂)_d-R¹³, substituted
 C₁₋₆alkyldisulfide, substituted phenyldisulfide, substituted C₁₋₆alkylurea,
 substituted phenylurea, substituted phenylthiourea or substituted
 C₁₋₆alkylthiourea wherein the substituents are selected from the group
 consisting of C₁₋₆alkyl, haloC₁₋₆alkyl, halogen, hydroxyl, carboxylic acid,
 sulfonic acid, phosphonic acid, amine, amidine, acetamide, and nitrile;

where

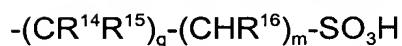
d is 0-8;

R¹³ is thioC₁₋₆alkylcarbonyl;

substituted C₁₋₆alkyl

where the alkyl substituents are selected from one or more members of the group consisting of hydroxyl, carboxylic acid, sulfonic acid, phosphonic acid, amine, amidine, acetamide, nitrile, thiol, C₁₋₆alkyldisulfide, C₁₋₆alkylsulfide, phenyldisulfide, urea, C₁₋₆alkylurea, phenylurea, thiourea, C₁₋₆alkylthiourea, phenylthiourea, substituted C₁₋₆alkyldisulfide, substituted phenyldisulfide, substituted C₁₋₆alkylurea, substituted phenylurea, substituted C₁₋₆alkylthiourea and substituted phenylthiourea

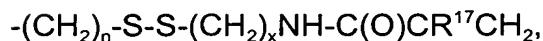
wherein the C₁₋₆alkyldisulfide, phenyldisulfide, C₁₋₆alkylurea, C₁₋₆alkylthiourea, phenylurea, and phenylthiourea substituents are selected from the group consisting of C₁₋₆alkyl, haloC₁₋₆alkyl, halogen, hydroxyl, carboxylic acid, sulfonic acid, phosphonic acid, amine, amidine, acetamide, and nitrile;



where R¹⁴, R¹⁵, and R¹⁶ are independently selected from the group consisting of hydrogen, halogen, hydroxyl, and C₁₋₆alkyl,

q is 1-6, and

m is 0-6;



where R¹⁷ is hydrogen or C₁₋₆alkyl,

n is 1-6, and

x is 1-6;



where R¹⁸, R¹⁹, and R²⁰ are independently selected from
the group consisting of hydrogen, halogen, hydroxyl, and
C₁₋₆alkyl,

t is 1-6, and

u is 0-6;

phenyl;

benzyl;

pyridinyl;

pyrimidinyl;

pyrazinyl;

benzimidazolyl;

benzothiazolyl;

benzotriazolyl;

naphthaloyl;

quinolinyl;

indolyl;

thiadiazolyl;

triazolyl;

4-methylpiperidin-1-yl;

4-methylpiperazin-1-yl;

substituted phenyl;

substituted benzyl;

substituted pyridinyl;

substituted pyrimidinyl;

substituted pyrazinyl;

substituted benzimidazolyl;

substituted benzothiazolyl;

substituted benzotriazolyl;

substituted naphthaloyl;

substituted quinolinyl;

substituted indolyl;

substituted thiadiazolyl;
substituted triazolyl;
substituted 4-methylpiperidin-1-yl; or
substituted 4-methylpiperazin-1-yl

5 wherein the substituents are selected from one or more
 members of the group consisting of C₁₋₆alkyl,
 haloC₁₋₆alkyl, halogen, sulfonic acid, phosphonic acid,
 hydroxyl, carboxylic acid, amine, amidine,
 N-(2-aminopyrimidine)sulfonyl,
10 N-(aminopyridine)sulfonyl, N-(aminopyrazine)sulfonyl,
 N-(2-aminopyrimidine)carbonyl,
 N-(aminopyridine)carbonyl, N-(aminopyrazine)carbonyl,
 N-(2-aminopyrimidine)phosphonyl,
 N-(2-aminopyridine)phosphonyl,
15 N-(aminopyrazine)phosphonyl,
 N-(aminobenzimidazolyl)sulfonyl,
 N-(aminobenzothiazolyl)sulfonyl,
 N-(aminobenzotriazolyl)sulfonyl,
 N-(aminoindolyl)sulfonyl, N-(aminothiazolyl)sulfonyl,
20 N-(aminotriazolyl)sulfonyl,
 N-(amino-4-methylpiperidinyl)sulfonyl,
 N-(amino-4-methylpiperazinyl)sulfonyl,
 N-(aminobenzimidazolyl)carbonyl,
 N-(aminobenzothiazolyl)carbonyl,
25 N-(aminobenzotriazolyl)carbonyl,
 N-(aminoindolyl)carbonyl, N-(aminothiazolyl)carbonyl,
 N-(aminotriazolyl)carbonyl,
 N-(amino-4-methylpiperidinyl)carbonyl,
 N-(amino-4-methylpiperazinyl)carbonyl,
30 N-(2-aminobenzimidazolyl)phosphonyl,
 N-(2-aminobenzothiazolyl)phosphonyl,

N-(2-aminobenzotriazolyl)phosphonyl,
 N-(2-aminoindolyl)phosphonyl,
 N-(2-aminothiazolyl)phosphonyl,
 N-(2-aminotriazolyl)phosphonyl,
 N-(amino-4-methylpiperidiny) phosphonyl,
 N-(amino-4-methylpiperazinyl) phosphonyl, acetamide,
 nitrile, thiol, C₁₋₆alkyldisulfide, C₁₋₆alkylsulfide, phenyl
 disulfide, urea, C₁₋₆alkylurea, phenylurea, thiourea,
 C₁₋₆alkylthiourea, phenylthiourea, substituted
 C₁₋₆alkyldisulfide, substituted phenyldisulfide, substituted
 C₁₋₆alkylurea, substituted C₁₋₆alkylthiourea, substituted
 phenylurea, and substituted phenylthiourea

wherein the C₁₋₆alkyldisulfide, phenyldisulfide,
 C₁₋₆alkylurea, C₁₋₆alkylthiourea, phenylurea, and
 phenylthiourea substituents are selected from the
 group consisting of C₁₋₆alkyl, haloC₁₋₆alkyl, halogen,
 hydroxyl, carboxylic acid, sulfonic acid, phosphonic
 acid, amine, amidine, acetamide, and nitrile;

b is 1-5;

p is 1-5;

R²¹ is hydrogen;

R²² is hydroxyl, sulfonic acid, phosphonic acid, carboxylic acid,
 thioC₁₋₆alkylcarbonyl, thioC₁₋₆alkylaminocarbonyl, C₁₋₆alkyldisulfide,
 phenyldisulfide, -C(O)NH(CH₂)₁₋₆-SO₃H, -C(O)NH(CH₂)₁₋₆-P(O)(OH)₂,
 -OR²³, -NH-R²³, -C(O)NH-(CH₂)_d-R²³, -S-(CH₂)_d-R²³, -(CH₂)_d-R²³, urea,
 C₁₋₆alkylurea, phenylurea, thiourea, C₁₋₆alkylthiourea, phenylthiourea,
 substituted C₁₋₆alkyldisulfide, substituted phenyldisulfide, substituted
 C₁₋₆alkylurea, substituted, C₁₋₆alkylthiourea substituted phenylurea or
 substituted phenylthiourea wherein the substituents are selected from
 the group consisting of C₁₋₆alkyl, haloC₁₋₆alkyl, halogen, hydroxyl,

carboxylic acid, sulfonic acid, phosphonic acid, amine, amidine, acetamide, and nitrile,

where

d is 0-8;

R²³ is thioC₁₋₆alkylcarbonyl,

C₁₋₆alkyl,

substituted C₁₋₆alkyl

where the alkyl substituents are selected from one or more members of the group consisting of C₁₋₆alkyl, halo C₁₋₆alkyl, halogen, hydroxyl, carboxylic acid, sulfonic acid, phosphonic acid, amine, amidine, acetamide, nitrile, thiol, C₁₋₆alkyldisulfide, C₁₋₆alkylsulfide, phenyldisulfide, urea, C₁₋₆alkylurea, phenylurea, thiourea, C₁₋₆alkylthiourea, phenylthiourea, substituted C₁₋₆alkyldisulfide, substituted phenyldisulfide, substituted C₁₋₆alkylurea, substituted phenylurea, substituted C₁₋₆alkylthiourea, and substituted phenylthiourea

wherein the C₁₋₆alkyldisulfide, phenyldisulfide, C₁₋₆alkylurea, C₁₋₆alkylthiourea, phenylurea, and phenylthiourea substituents are selected from the group consisting of C₁₋₆alkyl, haloC₁₋₆alkyl, halogen, hydroxyl, carboxylic acid, sulfonic acid, phosphonic acid, amine, amidine, acetamide, and nitrile;

$-(CR^{24}R^{25})_q-(CHR^{26})_m-SO_3H$

where R²⁴, R²⁵, and R²⁶ are independently selected from the group consisting of hydrogen, halogen, hydroxyl, and C₁₋₆alkyl,

q is 1-6, and

m is 0-6

$-(CH_2)_n-S-S-(CH_2)_x-NH-C(O)CR^{27}CH_2$,

where R²⁷ is hydrogen or C₁₋₆alkyl,

n is 1-6, and

x is 1-6;



where R^{28} , R^{29} , and R^{30} are independently selected from the group consisting of hydrogen, halogen, hydroxyl, and C_{1-6} alkyl,

t is 1-6, and

u is 0-6;

phenyl;

benzyl;

pyridinyl;

pyrimidinyl;

pyrazinyl;

benzimidazolyl;

benzothiazolyl;

benzotriazolyl;

naphthaloyl;

quinolinyl;

indolyl;

thiadiazolyl;

triazolyl;

4-methylpiperidin-1-yl;

4-methylpiperazin-1-yl;

substituted phenyl;

substituted benzyl;

substituted pyridinyl;

substituted pyrimidinyl;

substituted pyrazinyl;

substituted benzimidazolyl;

substituted benzothiazolyl;

substituted benzotriazolyl;

substituted naphthaloyl;
substituted quinolinyll;
substituted indolyl;
substituted thiadiazolyl;
substituted triazolyl;
substituted 4-methylpiperidin-1-yl; or
substituted 4-methylpiperazin-1-yl,

wherein the substituents are selected from one or more
members of the group consisting of C₁₋₆alkyl,
haloC₁₋₆alkyl, halogen, sulfonic acid, phosphonic acid,
hydroxyl, carboxylic acid, amine, amidine,
N-(2-aminopyrimidine)sulfonyl,
N-(aminopyridine)sulfonyl, N-(aminopyrazine)sulfonyl,
N-(2-aminopyrimidine)carbonyl,
N-(aminopyridine)carbonyl, N-(aminopyrazine)carbonyl,
N-(2-aminopyrimidine)phosphonyl,
N-(2-aminopyridine)phosphonyl,
N-(aminopyrazine)phosphonyl,
N-(aminobenzimidazolyl)sulfonyl,
N-(aminobenzothiazolyl)sulfonyl,
N-(aminobenzotriazolyl)sulfonyl,
N-(aminoindolyl)sulfonyl, N-(aminothiazolyl)sulfonyl,
N-(aminotriazolyl)sulfonyl,
N-(amino-4-methylpiperidinyl)sulfonyl,
N-(amino-4-methylpiperazinyl)sulfonyl,
N-(aminobenzimidazolyl)carbonyl,
N-(aminobenzothiazolyl)carbonyl,
N-(aminobenzotriazolyl)carbonyl,
N-(aminoindolyl)carbonyl, N-(aminothiazolyl)carbonyl,
N-(aminotriazolyl)carbonyl,
N-(amino-4-methylpiperidinyl)carbonyl,

N-(amino-4-methylpiperazinyl)carbonyl,
 N-(2-aminobenzimidazolyl)phosphonyl,
 N-(2-aminobenzothiazolyl)phosphonyl,
 N-(2-aminobenzotriazolyl)phosphonyl,
 N-(2-aminoindolyl)phosphonyl,
 N-(2-aminothiazolyl)phosphonyl,
 N-(2-aminotriazolyl)phosphonyl,
 N-(amino-4-methylpiperidinyl) phosphonyl,
 N-(amino-4-methylpiperazinyl) phosphonyl, acetamide,
 nitrile, thiol, C₁₋₆alkyldisulfide, C₁₋₆alkylsulfide, phenyl
 disulfide, urea, C₁₋₆alkylurea, phenylurea, thiourea,
 C₁₋₆alkylthiourea, phenylthiourea, substituted
 C₁₋₆alkyldisulfide, substituted phenyldisulfide, substituted
 C₁₋₆alkylurea, substituted C₁₋₆alkylthiourea, substituted
 phenylurea, and substituted phenylthiourea
 wherein the C₁₋₆alkyldisulfide, phenyldisulfide,
 C₁₋₆alkylurea, C₁₋₆alkylthiourea, phenylurea, and
 phenylthiourea substituents are selected from the
 group consisting of C₁₋₆alkyl, haloC₁₋₆alkyl, halogen,
 hydroxyl, carboxylic acid, sulfonic acid, phosphonic
 acid, amine, amidine, acetamide, and nitrile;

w is 0-1;

Y is oxygen or sulfur;

R³¹ is hydrogen or C₁₋₆alkyl;

R³² is hydroxyl, sulfonic acid, phosphonic acid, carboxylic acid,
 thioC₁₋₆alkylcarbonyl, thioC₁₋₆alkylaminocarbonyl, -C(O)NH-(CH₂)_d-R³³,
 -O-R³³, -NH-R³³, -S-(CH₂)_d-R³³, -(CH₂)_d-R³³, C₁₋₆alkyldisulfide,
 phenyldisulfide, urea, C₁₋₆alkylurea, phenylurea, thiourea,
 C₁₋₆alkylthiourea, phenylthiourea, C₁₋₆alkylamine, phenylamine,
 substituted C₁₋₆alkyldisulfide, substituted phenyldisulfide, substituted
 phenylurea, substituted C₁₋₆alkylamine, substituted phenylamine,

substituted phenylthiourea, substituted C₁₋₆alkylurea or substituted C₁₋₆alkylthiourea wherein the substituents are selected from the group consisting of C₁₋₆alkyl, haloC₁₋₆alkyl, halogen, hydroxyl, carboxylic acid, sulfonic acid, phosphonic acid, amine, amidine, acetamide, and nitrile

where

d is 0-8;

R³³ is thioC₁₋₆alkylcarbonyl,

C₁₋₆alkyl,

substituted C₁₋₆alkyl

where the alkyl substituents are selected from one or more members of the group consisting of

C₁₋₆alkyl, halo C₁₋₆alkyl, halogen, hydroxyl, carboxylic acid, sulfonic acid, phosphonic acid, amine, amidine, acetamide, nitrile, thiol,

C₁₋₆alkyldisulfide, C₁₋₆alkylsulfide, phenyldisulfide, urea, C₁₋₆alkylurea, phenylurea, thiourea,

C₁₋₆alkylthiourea, phenylthiourea, substituted C₁₋₆alkyldisulfide, substituted phenyldisulfide,

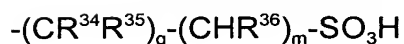
substituted C₁₋₆alkylurea, substituted phenylurea, substituted C₁₋₆alkylthiourea or substituted phenylthiourea

wherein the C₁₋₆alkyldisulfide, phenyldisulfide,

C₁₋₆alkylurea, C₁₋₆alkylthiourea, phenylurea, and

phenylthiourea substituents are selected from the

group consisting of C₁₋₆alkyl, haloC₁₋₆alkyl, halogen, hydroxyl, carboxylic acid, sulfonic acid, phosphonic acid, amine, amidine, acetamide, and nitrile;



where R³⁴, R³⁵, and R³⁶ are independently selected from the group consisting of hydrogen, halogen, hydroxyl, and C₁₋₆alkyl,

q is 1-6, and

m is 0-6;

$-(CH_2)_n-S-S-(CH_2)_x-NH-C(O)CR^{37}CH_2$,

where R^{37} is hydrogen or C_{1-6} alkyl,

n is 1-6, and

x is 1-6;

$-(CR^{38}R^{39})_t-(CHR^{40})_u-P(O)(OH)_2$

where R^{38} , R^{39} , and R^{40} are independently selected from the group consisting of hydrogen, halogen, hydroxyl, and C_{1-6} alkyl,

t is 1-6, and

u is 0-6;

phenyl;

benzyl;

pyridinyl;

pyrimidinyl;

pyrazinyl;

benzimidazolyl;

benzothiazolyl;

benzotriazolyl;

naphthaloyl;

quinolinyl;

indolyl;

thiadiazolyl;

triazolyl;

4-methylpiperidin-1-yl;

4-methylpiperazin-1-yl;

substituted phenyl;

substituted benzyl;

substituted pyridinyl;

substituted pyrimidinyl;

substituted pyrazinyl;
substituted benzimidazolyl;
substituted benzothiazolyl;
substituted benzotriazolyl;
5 substituted naphthaloyl;
substituted quinolinyll;
substituted indolyl;
substituted thiadiazolyl;
substituted triazolyl;
10 substituted 4-methylpiperidin-1-yl; or
substituted 4-methylpiperazin-1-yl,

wherein the substituents are selected from one or more
members of the group consisting of C₁₋₆alkyl,
haloC₁₋₆alkyl, halogen, sulfonic acid, phosphonic acid,
15 hydroxyl, carboxylic acid, amine, amidine,
N-(2-aminopyrimidine)sulfonyl,
N-(aminopyridine)sulfonyl, N-(aminopyrazine)sulfonyl,
N-(2-aminopyrimidine)carbonyl,
N-(aminopyridine)carbonyl, N-(aminopyrazine)carbonyl,
20 N-(2-aminopyrimidine)phosphonyl,
N-(2-aminopyridine)phosphonyl,
N-(aminopyrazine)phosphonyl,
N-(aminobenzimidazolyl)sulfonyl,
N-(aminobenzothiazolyl)sulfonyl,
25 N-(aminobenzotriazolyl)sulfonyl,
N-(aminoindolyl)sulfonyl, N-(aminothiazolyl)sulfonyl,
N-(aminotriazolyl)sulfonyl,
N-(amino-4-methylpiperidinyl)sulfonyl,
N-(amino-4-methylpiperazinyl)sulfonyl,
30 N-(aminobenzimidazolyl)carbonyl,
N-(aminobenzothiazolyl)carbonyl,

N-(aminobenzotriazolyl)carbonyl,
 N-(aminoindolyl)carbonyl, N-(aminothiazolyl)carbonyl,
 N-(aminotriazolyl)carbonyl,
 N-(amino-4-methylpiperidinyl)carbonyl,
 N-(amino-4-methylpiperazinyl)carbonyl,
 N-(2-aminobenzimidazolyl)phosphonyl,
 N-(2-aminobenzothiazolyl)phosphonyl,
 N-(2-aminobenzotriazolyl)phosphonyl,
 N-(2-aminoindolyl)phosphonyl,
 N-(2-aminothiazolyl)phosphonyl,
 N-(2-aminotriazolyl)phosphonyl,
 N-(amino-4-methylpiperidinyl) phosphonyl,
 N-(amino-4-methylpiperazinyl) phosphonyl, acetamide,
 nitrile, thiol, C₁₋₆alkyldisulfide, C₁₋₆alkylsulfide, phenyl
 disulfide, urea, C₁₋₆alkylurea, phenylurea, thiourea,
 C₁₋₆alkylthiourea, phenylthiourea, substituted
 C₁₋₆alkyldisulfide, substituted phenyldisulfide, substituted
 C₁₋₆alkylurea, substituted C₁₋₆alkylthiourea, substituted
 phenylurea, and substituted phenylthiourea

wherein the C₁₋₆alkyldisulfide, phenyldisulfide,
 C₁₋₆alkylurea, C₁₋₆alkylthiourea, phenylurea, and
 phenylthiourea substituents are selected from the
 group consisting of C₁₋₆alkyl, haloC₁₋₆alkyl, halogen,
 hydroxyl, carboxylic acid, sulfonic acid, phosphonic
 acid, amine, amidine, acetamide, and nitrile;

R⁴¹ is hydrogen, C₁₋₆alkyl, phenyl, C₁₋₆alkylcarbonyl, phenylcarbonyl,
 substituted C₁₋₆alkyl, substituted phenyl, substituted C₁₋₆alkylcarbonyl or
 substituted phenylcarbonyl,

wherein

the substituents are selected from the group consisting of
 C₁₋₆alkyl, haloC₁₋₆alkyl, halogen, hydroxyl, carboxylic acid,

sulfonic acid, phosphonic acid, amine, amidine, acetamide, and nitrile.

As used herein, the term "lens" refers to ophthalmic devices that reside in or on the eye. These devices can provide optical correction or may be cosmetic. The term lens includes but is not limited to soft contact lenses, hard contact lenses, intraocular lenses, overlay lenses, ocular inserts, and optical inserts. Soft contact lenses are made from silicone elastomers or hydrogels, which include but are not limited to silicone hydrogels and fluorohydrogels. These hydrogels contain hydrophobic and/or hydrophilic monomers that are covalently bound to one another in the cured lens. As used herein the term "polymers" means copolymers, homopolymers, or mixtures thereof. The monomers of Formula I, II, III or IV, or their homopolymers, are added to the monomer mix of contact lenses, prior to polymerization in an amount based on the weight percent of the initial monomer mix, including a suitable diluent if said diluent is used in the preparation of the polymer. The weight percentage of the monomers of the invention can vary with the lens formulation. The maximum percentage of monomers of Formula I, II, III or IV is the percentage that does not compromise the physical properties of the resulting contact lens, such as, but not limited to modulus, of the resulting lens. The minimum percentage of monomers of Formula I, II, III or IV is an amount that allows the incorporation of a sufficient amount of silver into a lens. Preferably, about 0.01 to about 20.0 weight percent of monomers of Formula I, II, III or IV are added, to a contact lens formulation, more preferably, about 0.01 to about 1.5 weight percent, even more preferably, about 0.01 to about 0.4 weight percent, most preferably, about 0.2 weight percent.

Monomers of Formula I, II, III or IV are added to the soft contact lens formulations described in U.S. Pat. No. 5,710,302, WO 9421698, EP 406161, JP 2000016905, U.S. Pat. No. 5,998,498, US Pat. App. No. 09/532,943 and U.S. Pat. No. 6,087,415. In addition, monomers of Formula I, II, III or IV may be added to the formulations of commercial soft contact lenses. Examples of

commercially available soft contact lenses formulations include but are not limited to, the formulations of etafilcon A, genfilcon A, lenefilcon A, polymacon, acquafilcon A, balafilcon A, and lotrafilcon A. The preferable contact lens formulations are etafilcon A, balafilcon A, and silicone hydrogels, as prepared in U.S. Pat. No. 5,760,100; U.S. Pat. No. 5,776,999; U.S. Pat. No. 5,849,811; U.S. Pat. No. 5,789,461; U.S. Pat. No. 5,998,498, US Pat. App. No. 09/532,943, a continuation-in-part of U.S. Pat. App. No. 09/532,943, filed on August 30, 2000, and U.S. Pat. No. 6,087,415. These patents are hereby incorporated by reference for the hydrogel compositions contained therein. Lenses prepared from the aforementioned formulations and the monomers of Formula II, II, III or IV may be coated with a number of agents that are used to coat lenses. For example, the procedures, compositions, and methods of U.S. Pat. Nos. 3,854,982; 3,916,033; 4,920,184; and 5,002,794; 5,712,327; and 6,087,415 as well as WO 0127662, may be used and these patents are hereby incorporated by reference for those procedures, compositions, and methods. In addition to the cited coating patents, there are other methods of treating a lens once it is formed. The lenses of this invention may be treated by these methods and the following publications which illustrate these methods are hereby incorporated by reference in their entirety, U. S. Pat. No. 5,453,467; U.S. Pat. No. 5,422,402; WO 9300391; U.S. Pat. No. 4,973,493; and U.S. No. Pat 5,350,800.

Hard contact lenses are made from polymers that include but are not limited to polymers of poly(methyl)methacrylate, silicon acrylates, fluoroacrylates, fluoroethers, polyacetylenes, and polyimides, where the preparation of representative examples may be found in JP 200010055; JP 6123860; and U.S. Pat. No. 4,330,383. Intraocular lenses of the invention can be formed using known materials. For example, the lenses may be made from a rigid material including, without limitation, polymethyl methacrylate, polystyrene, polycarbonate, or the like, and combinations thereof. Additionally, flexible materials may be used including, without limitation, hydrogels, silicone materials, acrylic materials, fluorocarbon materials and the like, or combinations thereof. Typical intraocular lenses are described in WO 0026698; WO 0022460;

WO 9929750; WO 9927978; WO 0022459; and JP 2000107277. The polymerizable monomers of Formula I, II, III or IV may be added to hard contact lens formulations and intraocular lens formulations in the same manner and at the same percentage as described above for soft contact lenses. All of the references mentioned in this application are hereby incorporated by reference in their entirety.

As used herein, the term "silver" refers to silver metal that is incorporated into a lens. While not wanting to be bound as to the oxidation state of the silver (Ag^0 , Ag^{1+} , or Ag^{2+}), that is incorporated into the lens, silver may be added to the lens by washing the cured and hydrated lens in a silver solution such as silver nitrate in deionized water ("DI"). Other sources of silver include but are not limited to silver acetate, silver citrate, silver iodide, silver lactate, silver picrate, and silver sulfate. The concentration of silver in these solutions can vary from the concentration required to add a known quantity of silver to a lens to a saturated silver solution. In order to calculate the concentration of the silver solution needed, the following calculation is used: the concentration of silver solution is equal to the desired amount of silver per lens, multiplied by the dry weight of the lens divided by the total volume of treating solution.

$$\text{silver solution concentration } (\mu\text{g/mL}) = [\text{desired silver in lens } (\mu\text{g/g}) \times \text{average dry lens weight (g)}] / \text{total volume of treating solution (mL)}$$

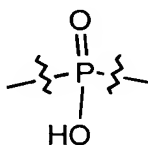
For example, if one requires a lens containing 40 $\mu\text{g/g}$ of silver, the dry weight of the lens is 0.02 g, and the vessel used to treat said lens has a volume of 3mL, the required silver concentration would be 0.27 $\mu\text{g/mL}$.

Silver solutions containing anywhere from about 0.10 $\mu\text{g/mL}$ to 0.3 grams/mL have been used to prepare the lenses of the invention. Aside from deionized water, other liquid mediums can be used such as water, aqueous buffered solutions and organic solutions such as polyethers or alcohols. Typically, the lens is washed in the silver solution for about 60 minutes, though the time may vary from about 1 minute to about 2 hours and at temperatures ranging from about 5°C to about 130°C. After the silver treatment the lenses are washed with several portions of water to obtain a lens where silver is

incorporated into the polymer. The amount of silver that is incorporated into the lenses ranges from about 20 ppm to about 100,000 ppm, where any lens containing at least about 20 ppm has antimicrobial properties. The preferred amount of silver that is incorporated into the lens is about 20 ppm to about 4,000 ppm, more preferably, 20 ppm to about 1,500 ppm, even more preferably about 30 ppm to about 600 ppm, and most preferably about 30 ppm to about 75 ppm.

The term "antimicrobial" refers to a lens that exhibit one or more of the following properties - the inhibition of the adhesion of bacteria or other microbes to the lenses, the inhibition of the growth of bacteria or other microbes on the lenses, and the killing of bacteria or other microbes on the surface of the lenses or in a radius extending from the lenses (hereinafter adhesion of bacteria or other microbes to the lenses, the growth of bacteria or other microbes to the lenses and the presence of bacterial or other microbes on the surface of lenses is collectively referred to as "microbial production"). The lenses of the invention inhibit the microbial production by at least 25%. Preferably, the lenses of the invention exhibit at least a 1-log reduction ($\geq 90\%$ inhibition) of viable bacteria or other microbes, more preferably a 2-log reduction ($\geq 99\%$ inhibition) of viable bacteria or other microbes. Such bacteria or other microbes include but are not limited to those organisms found in the eye, particularly *Pseudomonas aeruginosa*, *Acanthamoeba species*, *Staphylococcus aureus*, *E. coli*, *Staphylococcus epidermidis*, and *Serratia marcesens*. Preferably, said antimicrobial lens is a clear lens, that has clarity comparable to currently available commercial lenses such as but not limited to, etafilcon A, genfilcon A, lenefilcon A, polymacon, aquafilcon A, balafilcon A, and lotrafilcon A.

The term "phosphonyl" refers to a radical having the following structure



With respect to the monomers of Formula I, there are some monomers that are preferred. The preferred monomers of Formula I include monomers where R^1 is hydrogen or C_{1-3} alkyl;

R² is NH-R³;

d is 0

R³ is substituted phenyl, -(CR⁴ R⁵)_q-(CHR⁶)_m-SO₃H, -(CR⁸R⁹)_t-(CHR¹⁰)_u-P(O)(OH)₂,
or -(CH₂)_n-S-S-(CH₂)_xNH-C(O)CR⁷CH₂;

5 R⁴ is hydrogen or C₁₋₃alkyl;

R⁵ is hydrogen or C₁₋₃alkyl;

R⁶ is hydrogen or C₁₋₃alkyl;

q is 1-3;

m is 1-3;

10 R⁷ is hydrogen or C₁₋₃alkyl;

R⁸ is hydrogen or C₁₋₃alkyl;

R⁹ is hydrogen or C₁₋₃alkyl;

R¹⁰ is hydrogen or C₁₋₃alkyl;

t is 1-3;

15 u is 1-3;

n is 2-4; and

x is 2-4.

The more preferred monomers of Formula I include monomer where

R¹ is hydrogen or methyl;

20 R² is NH-R³;

R³ is -(CR⁴ R⁵)_q-(CHR⁶)_m-SO₃H, -(CR⁸R⁹)_t-(CHR¹⁰)_u-P(O)(OH)₂ or
-(CH₂)_n-S-S-(CH₂)_xNH-C(O)CHR⁷CH₂;

R⁴ is hydrogen or methyl;

R⁵ is hydrogen or methyl;

25 q is 1-2;

m is 1-2;

R⁶ is hydrogen or methyl;

R⁷ is hydrogen;

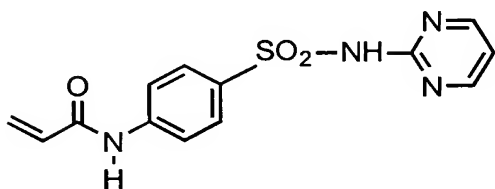
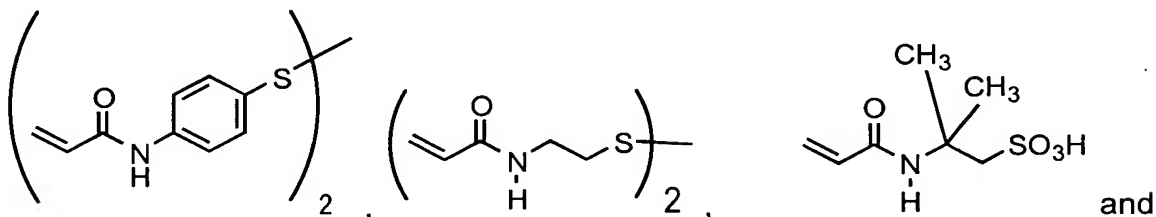
R⁸ is hydrogen or methyl;

30 R⁹ is hydrogen or methyl;

R¹⁰ is hydrogen or methyl;

t is 1;
u is 1-2;
n is 2-3; and
x is 2-3.

5 The most preferred monomers of Formula I include the following monomers



10 With respect to the monomers of Formula II, there are some monomers that are preferred. The preferred monomers of Formula II include monomers where

a is 1-2;

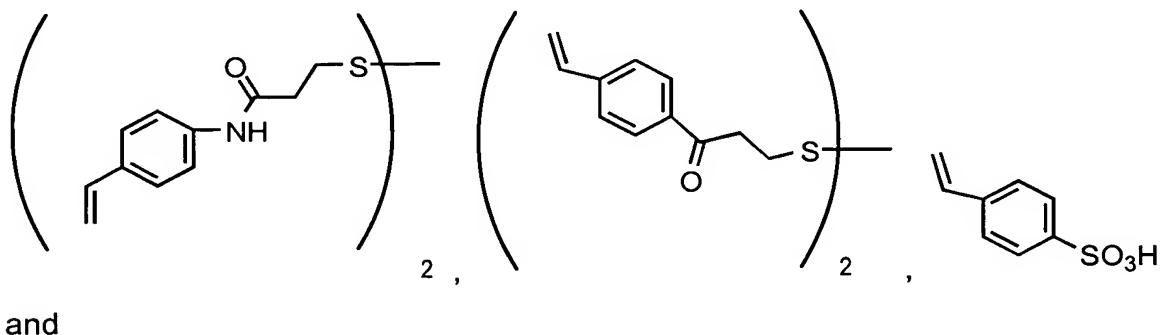
R¹¹ is hydrogen or C₁₋₃alkyl;

R¹² is sulfonic acid, carboxylic acid, phosphonic acid, C₁₋₆alkyldisulfide,

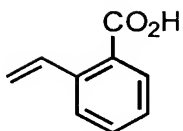
15 C₁₋₆alkylsulfide, phenyldisulfide, substituted phenyldisulfide or NH-R¹³;

R¹³ is thioC₁₋₆alkylcarbonyl.

The most preferred monomers of Formula II include the following monomers



20



With respect to the monomers of Formula III, there are some monomers that are preferred. The preferred monomers of Formula III include monomers where

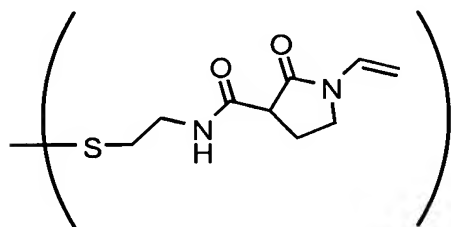
p is 1-3;

b is 1-2;

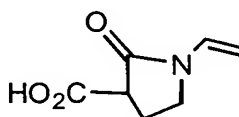
R²¹ is hydrogen;

R²² is sulfonic acid, phosphonic acid, carboxylic acid, thioC₁₋₆alkylcarbonyl, thioC₁₋₆alkylaminocarbonyl, C₁₋₆alkyldisulfide, C₁₋₆alkylsulfide, phenyldisulfide, substituted phenyldisulfide, H₃OS-(CH₂)₁₋₆NHC(O) or (HO)₂(O)P-(CH₂)₁₋₆NHC(O)-.

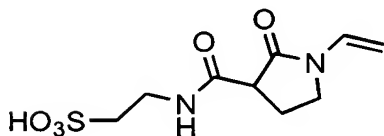
The most preferred monomers of Formula III include the following monomers



2 ,



and



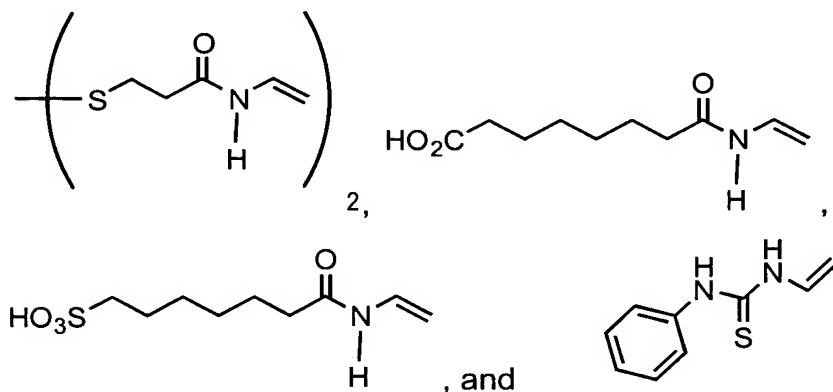
With respect to the monomers of Formula IV, there are some monomers that are preferred. The preferred monomers of Formula IV include monomers where

w is 0-1;

R³¹ is hydrogen;

R³² is amine, C₁₋₃alkylamine, phenylamine, substituted phenylamine,
thioC₁₋₃alkylcarbonyl;
R⁴¹ is hydrogen

- 5 The most preferred monomers of Formula IV include the following monomers

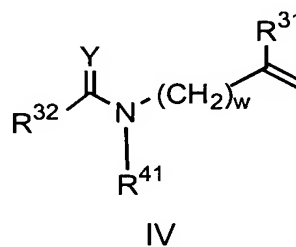
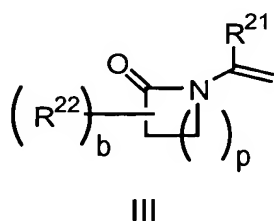
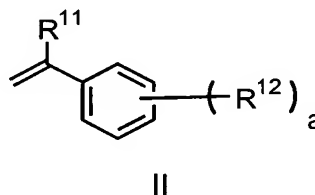
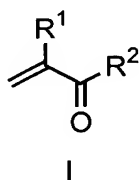


For monomers of all formula, the preferred points of attachment are as follows pyridinyl is 4, for pyrimidinyl is 2, for benzimidazolyl is 2, for
10 benzothiazole is 2, for benzotriazolyl is 5, for quinolinyl is 2, for indolyl is 4 or 5, for thiadiazole is 3, or 5, and for triazolyl is 3 or 5, where positions containing hydrogen may be substituted with the named substituents.

Further, the invention includes an antimicrobial lens comprising,
consisting essentially of, or consisting of, silver and a polymer comprising a
15 binding monomer where said cured lens can reversibly bind silver. The terms antimicrobial, lens, and silver all have their aforementioned meanings and preferred ranges. The term "binding monomer" means any polymerizable monomer that can reversibly bind silver. The term "cured lens" refers to contact lens monomer formulations polymerized with binding monomers. The potential
20 ability of a cured lens to reversibly bind silver can be estimated by examining the stability constant of the selected binding monomers. These estimates can be determined by known methods. (See R.I. Tilley, Aust J. Chem. 1990, 43,1573). The log of the stability constants, β_n , of the cured lense of the invention are about 0.6 to about 15.0; that is, $\log \beta_n = [AgL_n^+]/([Ag^+][L]^n)$ where β_n = stability
25 constant for a cured lens (L) that binds n molecules of silver (Ag^+), preferably, about 2 to about 7.3, and more preferably about 3.6 to 6.9.

The advantages of the antimicrobial lenses of the invention are many. For example, other antimicrobial lenses that incorporate silver usually contain silver coordinated to some inorganic particulate matter (see US Pat. No. 5,213,801, discussing the use of silver ceramics). Often that particulate matter is visible to the naked or magnified eye, and it can affect the visual acuity of the user. However, the lenses of the invention do not have this problem. The monomers of Formula I, II, III or IV and other binding monomers are generally soluble with all of the other components of the antimicrobial lenses. Therefore when the lenses are produced they do not have substantial particulate matter due to their antimicrobial components. The antimicrobial lenses of the invention have comparable clarity to commercial lenses such as etafilcon A, genfilcon A, lenefilcon A, polymacon, aquafilcon A, balafilcon A, and lotrafilcon A.

Further, the invention includes a method of producing an antimicrobial lens comprising, silver and a polymer comprising a monomer of Formula I, II, III or IV



wherein $\text{R}^1 - \text{R}^{41}$, Y, a, q, m, n, p, d, b, t, u, w, and x are as described above wherein

the method comprises, consists essentially of, or consists of the steps of

- (a) preparing a lens comprising a monomer of Formula I, II, III or IV and
- (b) treating said lens with a silver solution.

The terms lens, antimicrobial, lens, silver, $R^1 - R^{41}$, Y, a, q, m, n, p, d, b, t, u, w, and x, all have their aforementioned meanings and preferred ranges. The term, "silver solution" refers to any liquid medium containing silver. The liquid medium includes but is not limited to water, deionized water, aqueous buffered solutions, alcohols, polyols, and glycols, where the preferred medium is deionized water.

The silver of the solution is typically a silver salt such as silver nitrate, silver acetate, silver citrate, silver iodide, silver lactate, silver picrate, and silver sulfate.

The concentration of silver in these solutions can vary from the concentration required to add a known quantity of silver to a lens to a saturated silver solution.

In order to calculate the concentration of the silver solution needed, the following calculation is used: the concentration of silver solution is equal to the desired amount of silver per lens, multiplied by the dry weight of the lens divided by the total volume of treating solution.

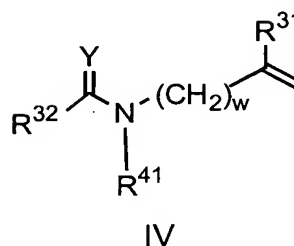
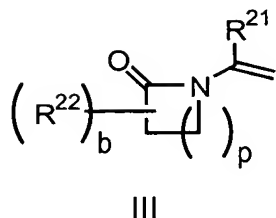
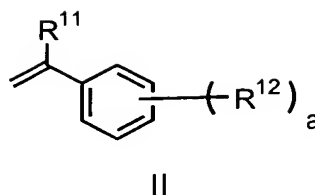
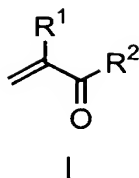
$$\text{silver solution concentration } (\mu\text{g/mL}) = [\text{desired silver in lens } (\mu\text{g/g}) \times \text{average dry lens weight (g)}] / \text{total volume of treating solution (mL)}$$

For example, if one requires a lens containing 40 $\mu\text{g/g}$ of silver, the dry weight of the lens is 0.02g, and the vessel used to treat said lens has a volume of 3mL, the required silver concentration would be 0.27 $\mu\text{g/mL}$.

Silver solutions containing anywhere from about 0.10 $\mu\text{g/mL}$ to 0.3 grams/mL have been used to prepare the lenses of the invention. Aside from deionized water, other liquid mediums can be used such as water, aqueous buffered solutions and organic solutions such as polyethers, or alcohols.

Typically, the lens is washed in the silver solution for about 60 minutes, though the time may vary from about 1 minute to about 2 hours and at temperatures ranging from about 5°C to about 130°C. After the silver treatment the lenses are washed with several portions of water to obtain a lens where silver is incorporated into the polymer.

Still further, the invention includes a lens case comprising, consisting essentially of, or consisting of silver and a polymer of a monomer of Formula I, II, III or IV



5

wherein R¹ - R⁴¹, Y, a, q, m, n, p, d, b, t, u, w, and x are as described above

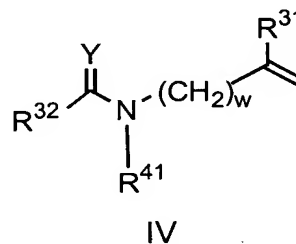
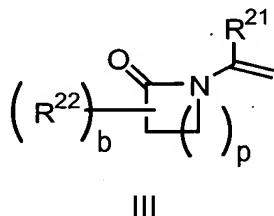
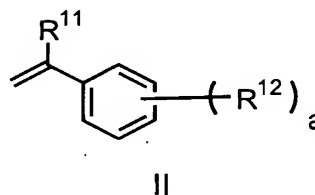
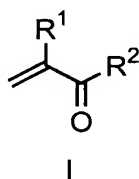
The terms lens, silver, R¹ - R⁴¹, Y, a, q, m, n, p, d, b, t, u, w, and x, all have their aforementioned meanings and preferred ranges. The term lens case refers to a container that is adapted to define a space in which to hold a lens when that lens is not in use. This term includes packaging for lenses, where packaging includes any unit in which a lens is stored after curing. Examples of this packaging include but are not limited to single use blister packs, multiple use storage cases and the like.

One such container is illustrated in Figure 3 of U.S. Pat. 5,515,117 which is hereby incorporated by reference in its entirety. The polymers of Formula I, II, III or IV can be incorporated in the lens container 22, the cover 24, or the lens basket 26, where they are preferably incorporated into the lens container or the lens basket.

Aside from the polymers of Formula I, II, III or IV the container components may be made of a transparent, thermo-plastic polymeric material, such as polymethylmethacrylate, polyolefins, such as poly-ethylene, polypropylene, their copolymers and the like; polyesters, polyurethanes; acrylic polymers, such as polyacrylates and polymethacrylates; polycarbonates and the like and is made, or any combination thereof, e.g., molded, using conventional techniques as a single unit.

Silver may be incorporated into the lens container in the same manner that it is incorporated into the antimicrobial lenses of the invention. More specifically, the polymers of Formula I, II, III or IV are combined with the formulation of the other components, molded, cured, and subsequently treated with a silver solution. Preferably, polymers of Formula I, II, III or IV are present in any or all of the lens case components at about 0.01 to about 10.0 weight percent (based on the initial monomer mix), more preferably about 0.01 to about 1.5 percent. Storing lenses in such an environment inhibits the growth of bacteria on said lenses and adverse effects that are caused by the proliferation of bacterial. Another example of such a lens case is the lens case can be found in U.S. Pat. No. 6,029,808 which is hereby incorporated by reference for the blister pack housing for a contact lens disclosed therein.

Yet still further, the invention includes a method of reducing the adverse effects associated with microbial production in the eye of a mammal, comprising, consisting essentially of, or consisting of providing an antimicrobial lens wherein said lens comprises silver and a polymer of a monomer of Formula I, II, III or IV



wherein $R^1 - R^{41}$, Y, a, q, m, n, p, d, b, t, u, w, and x are as described above

The terms lens, antimicrobial, lens, silver, $R^1 - R^{41}$, Y, a, q, m, n, p, d, b, t, u, w and x, all have their aforementioned meanings and preferred ranges. The phrase "adverse effects associated with microbial production" includes but is not

limited to, ocular inflammation, contact lens related peripheral ulcers, contact lens associated red eye, infiltrative keratitis, and microbial keratitis.

Providing a lens that fits a wide range of patients has been a quest of eye care practitioners and lens manufactures for a number of years. In order to produce such a lens, many variables, such as lens material, design, surface treatments, and additional components such as ophthalmic drugs, tints, dyes and pigments can come into play. For example it has been shown that if one adds too much of an additional component, such as an antimicrobial agent, a lens that will become adhered to the eye is produced. However, if one is attempting to produce an antimicrobial lens, a balance should be struck between producing a lens that contains enough antimicrobial agent to produce the desired effect without producing a lens that adheres to the eye.

One way to assess if a lens fit is acceptable (i.e. the lens is not adhered) is to assess the tightness of the fit of a lens. (Young, G. et al., Influence of Soft Contact Lens Design on Clinical Performance, Optometry and Vision Science, Vol 70, No., 5 pp. 394-403) Tightness of a lens may be assessed using an *in vivo* push up test. In that test, a lens is placed on a patient's eye. Subsequently, an eye care practitioner presses his or her finger digitally upward against the lower lid of the patient's eye and observes whether the lens moves on the patient's eye (*Id.*). Lenses that do not move under these circumstances are not considered to be a good fit for the patient's eye, for lenses that are too tight will not move when the patient blinks and may become uncomfortable. Therefore one of the objects of this invention is to produce an antimicrobial lens that does not adhere to the patient's eye.

To meet this objective, the invention includes an antimicrobial lens comprising, consisting essentially of, or consisting of silver, wherein said lens has sufficient movement on the eye of a patient. The terms lens, antimicrobial, lens, silver, $R^1 - R^{41}$, Y, a, q, m, n, p, d, b, t, u, w, and x, all have their aforementioned meanings and preferred ranges. The phrase "movement on the eye of a patient" refers to whether a lens, when placed on the eye of a patient moves under the push-up test described above. This test is described in further

detail in Contact Lens Practice, Chapman & Hall, 1994, edited by M. Ruben and M. Guillon, pgs. 589-99. Under this test lenses are given an -2 rating if they do not move on the eye of a patient in the digital push-up test. Therefore lenses that score greater than a "-2" on the digital push-up test are lenses that move on a patient's eye. In a statistically significant patient population, lenses that may be suitable for one patient may not be suitable for another. Therefore, lenses having sufficient movement are lenses that move on at least about 50 to about 100% of a given patient population. Preferably, said lenses move on about 75 to about 100%, of patients, more preferably, about 80 to about 100%, most preferably about 90 to about 100%.

The lenses of the invention are one method of making lenses that contain silver and have sufficient movement on the eye of a patient; however, they are not only lenses containing silver that may have sufficient movement. Other methods of incorporating into contact lenses may be used, provided that those methods produce lenses having sufficient movement on the eye of a patient.

In order to illustrate the invention the following examples are included. These examples do not limit the invention. They are meant only to suggest a method of practicing the invention. Those knowledgeable in contact lenses as well as other specialties may find other methods of practicing the invention. However, those methods are deemed to be within the scope of this invention.

EXAMPLES

The following abbreviations were used in the examples

APDS = acrylamidophenylsulfide

AMPSA = 2-acrylamido-2-methyl-1-propanesulfonic acid;

CYST = N,N'-(bisacryloyl)cystamine;

PVP= polyvinylpyrrolidinone;

MAA = methacrylic acid;

PAA = poly(acrylic acid)

ATU = allylthiourea;

VIM = vinyl imidazole;

MABP = methacrylamido bipyrimidine;

MAHB = 4-methacryloxy-2-hydroxybenzophenone;

PSPM = N-[p-(N-pyrimidin-2-sulfamoyl)phenyl]methacrylamide

Cell/prot = (Acrylamidomethyl)cellulose acetate propionate

3M3P = 3-methyl-3-propanol

5 D3O = 3,7-dimethyl-3-octanol

TAA = t-amyl alcohol

BAGE = glycerin esterified with boric acid

DI= deionized water;

PBS = phosphate-buffered saline, pH 7.4 ± 0.2 ;

10 TPBS = Phosphate-buffered saline with 0.05% Tween™ 80, pH 7.4 ± 0.2 ;

TSA = sterile tryptic soy agar;

TSB = sterile tryptic soy broth;

60% IPA = isopropyl alcohol, 60% v/v DI;

70% IPA = isopropyl alcohol, 70% v/v DI;

15 10% IPA = isopropyl alcohol, 10% v/v DI;

MVD = modified vortex device;

TBACB = tetrabutyl ammonium-m-chlorobenzoate

TMI = dimethyl meta-isopropenyl benzyl isocyanate

MMA = methyl methacrylate

20 HEMA = hydroxyethyl methacrylate

Bloc-HEMA = 2-(trimethylsiloxy) ethyl methacrylate

TRIS = *tris*(trimethylsiloxy)-3-methacryloxypropylsilane

mPDMS = *mono*-methacryloxypropyl terminated polydimethylsiloxane MW = 800-1000

25 DMA = N,N-dimethylacrylamide

Blue HEMA = the reaction product of reactive blue number 4 and HEMA as described in Example 4 of U.S. Patent 5,944,853

DAROCUR 1173 = 2-hydroxy-2-methyl-1-phenyl-propan-1-one

EGDMA = ethyleneglycol dimethacrylate

30 TMPTMA = trimethyloyl propane trimethacrylate

TEGDMA = tetraethyleneglycol dimethacrylate

Norbloc = 2-(2'-hydroxy-5-methacryloxyethylphenyl)-2H-benzotriazole
CGI 1850 = 1:1 (w/w) blend of 1-hydroxycyclohexyl phenyl ketone and bis (2,6-dimethoxybenzoyl)-2,4,4-trimethylpentyl phosphine oxide

THF = tetrahydrofuran

5 HAM = hydroxyalkylmethacrylate, as described in US. Patent No. 5,98,498

w/w = weight/total weight

w/v = weight/total volume

v/v = volume/total volume

pHEMA = poly(hydroxyethyl) methacrylate coating as described in Example 14
10 of U.S. Serial No. 09/921,192, "Methods for Coating Articles by Mold Transfer"

The contact lenses of the invention were evaluated using the following biological assay, where Test B is the preferred method for determining inhibition of microbial production under the present invention.

Test A Inhibition of Bacterial Growth/Adhesion

15 A culture of *Pseudomonas aeruginosa*, ATCC# 15442 (ATCC, Rockville, MD) was grown overnight in a nutrient medium. The bacterial inoculum was prepared to result in a final concentration of approximately 1×10^8 colony forming units/mL. Three contact lenses were rinsed with phosphate buffered saline (PBS) pH 7.4 ± 0.2 . Each rinsed contact lens was combined with two (2)
20 mL of bacterial inoculum into a sterile glass vial, which was rotated in a shaker-incubator (100 rpm) for two (2) hrs. at $37 \pm 2^\circ\text{C}$. Each lens was rinsed with PBS to remove loosely bound cells, placed into 10 mL of PBS containing 0.05% w/v
25 TweenTM 80 and vortexed at 2000 rpm for three minutes. The resulting supernatant was enumerated for viable bacteria, and the results, reported of the detected viable bacteria attached to three lenses were averaged.

Test B Inhibition of Bacterial Growth/Adhesion

30 A culture of *Pseudomonas aeruginosa*, ATCC# 15442 (ATCC, Rockville, MD) was grown overnight in a nutrient medium. The bacterial inoculum was

prepared to result in a final concentration of approximately 1×10^6 colony forming units/mL. Three contact lenses were rinsed with phosphate buffered saline (PBS) pH 7.4 ± 0.2 . Each rinsed contact lens was combined with two (2) mL of bacterial inoculum into a sterile glass vial, which was rotated in a shaker-incubator (100 rpm) for 24 hr. at $35 \pm 2^\circ\text{C}$. Each lens was rinsed with PBS to remove loosely bound cells, placed into 10 mL of PBS containing 0.05% w/v Tween™ 80 and vortexed at 2000 rpm for three minutes. The resulting supernatant was enumerated for viable bacteria, and the results, reported of the detected viable bacteria attached to three lenses were averaged.

The formulations that were used to prepare the lenses of the invention were prepared as follows.

Macromer 2 Preparation

To a dry container housed in a dry box under nitrogen at ambient temperature was added 30.0 g (0.277 mol) of bis(dimethylamino)methylsilane, a solution of 13.75 mL of a 1M solution of TBACB (386.0 g TBACB in 1000 mL dry THF), 61.39 g (0.578 mol) of p-xylene, 154.28 g (1.541 mol) methyl methacrylate (1.4 equivalents relative to initiator), 1892.13 (9.352 mol) 2-(trimethylsiloxy)ethyl methacrylate (8.5 equivalents relative to initiator) and 4399.78 g (61.01 mol) of THF. To a dry, three-necked, round-bottomed flask equipped with a thermocouple and condenser, all connected to a nitrogen source, was charged the above mixture prepared in the dry box.

The reaction mixture was cooled to 15°C while stirring and purging with nitrogen. After the solution reached 15°C , 191.75 g (1.100 mol) of 1-trimethylsiloxy-1-methoxy-2-methylpropene (1 equivalent) was injected into the reaction vessel. The reaction was allowed to exotherm to approximately 62°C and then 30 mL of a 0.40 M solution of 154.4 g TBACB in 11 mL of dry THF was metered in throughout the remainder of the reaction. After the temperature of reaction reached 30°C and the metering began, a solution of 467.56 g (2.311

mol) 2-(trimethylsiloxy)ethyl methacrylate (2.1 equivalents relative to the initiator), 3636.6 g (3.463 mol) n-butyl monomethacryloxypropyl-polydimethylsiloxane (3.2 equivalents relative to the initiator), 3673.84 g (8.689 mol), TRIS (7.9 equivalents relative to the initiator) and 20.0 g

5 bis(dimethylamino)methylsilane was added.

The mixture was allowed to exotherm to approximately 38-42 °C and then allowed to cool to 30 °C. At that time, a solution of 10.0 g (0.076 mol) bis(dimethylamino)methylsilane, 154.26 g (1.541 mol) methyl methacrylate (1.4
10 equivalents relative to the initiator) and 1892.13 g (9.352 mol) 2-trimethylsiloxy)ethyl methacrylate (8.5 equivalents relative to the initiator) was added and the mixture again allowed to exotherm to approximately 40 °C. The reaction temperature dropped to approximately 30 °C and 2 gallons of THF were added to decrease the viscosity. A solution of 439.69 g water, 740.6 g methanol
15 and 8.8 g (0.068 mol) dichloroacetic acid was added and the mixture refluxed for 4.5 hours to de-block the protecting groups on the HEMA. Volatiles were then removed and toluene added to aid in removal of the water until a vapor temperature of 110 °C was reached.

20 The reaction flask was maintained at approximately 110 °C and a solution of 443 g (2.201 mol) TMI and 5.7 g (0.010 mol) dibutyltin dilaurate were added. The mixture was reacted until the isocyanate peak was gone by IR. The toluene was evaporated under reduced pressure to yield an off-white, anhydrous, waxy reactive monomer. The macromer was placed into acetone at a weight basis of
25 approximately 2:1 acetone to macromer. After 24 hrs, water was added to precipitate out the macromer and the macromer was filtered and dried using a vacuum oven between 45 and 60 °C for 20-30 hrs.

Macromer 1 Preparation

30 The procedure for Macromer 2 used except that 19.1 mole parts HEMA, 5.0 mole parts MAA, 2.8 mole parts MMA; 7.9 mole parts TRIS, 3.3, mole parts

mPDMS, and 2.0 mole parts TMI were used.

Macromer 3 Preparation

5 The procedure for Macromer 2 was used except that 19.1 mole parts HEMA, 7.9 mole parts TRIS, 3.3 mole parts mPDMS, and 2.0 mole parts TMI were used.

Macromer 4 Preparation

10 The procedure for Macromer 2 was used except that dibutyltin dilaurate was replaced with triethylamine.

Base Monomer Formations and Lens Preparation

15 Formulations A-R, listed in Table 1, are representative base monomer mixes (all amounts are calculated as weight percent of the total weight of the combination). The polymerizable monomers of the invention are added to these mixtures as indicated in Table 2 and contact lenses are prepared according to the following method.

20 Contact lenses are prepared by adding the indicated amount of the polymerizable monomer to about 10 g of the base monomer mix in the presence of 1 – 5%wt acetic acid (when Macromer 4 is used, no acetic acid is added) and a diluent suitable for compatibilizing the components, as indicated in Table 1. This mixture is sonicated at 25-37°C until all components are dissolved (30-120 minutes) and was subsequently loaded into an eight cavity lens mold of the type described in U.S. Pat. No. 4,640,489 and cured for 1200 sec at temperatures of, 25 but not limited to, 25 to 90°C, preferably between 45 to 75°C. Polymerization
25 occurred under a nitrogen purge and was either photoinitiated with 5 mW cm⁻² of UV light generated with an Andover Corp. 420PS10-25 AM39565-02 light filter, or photoinitiated with visible light generated with a Philips TL 20W/03T fluorescent bulb. The time of curing varied from 7 minutes to 60 minutes. After curing, the
30 molds are opened, and the lenses are either released in a 1:1 blend of water and ethanol, then leached in ethanol to remove any residual monomers and diluent, or

released in a 60% IPA/water, then leached in IPA/DI to remove any residual monomers and diluent. Finally the lenses were equilibrated in either physiological borate-buffered saline or de-ionized water.

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Table 1

Formulation	A	B	C	D	E	F	G	H	I	J	K	M	N	O	P	Q	R
Macromer	1	2	3	3	2	2	2	2	2	2	2	2	2	2	2	2	2
[Macromer]	30.00	25.00	60.00	20.00	17.98	17.98	18.00	19.98	17.98	17.98	19.98	40.00	18.00	18.00	18.00		
TRIS	0.00	18.00	0.00	40.00	21.00	21.00	14.00	8.00	20.00	25.00	20.00	20.00	14.00	14.00	14.00		
DMA	27.00	28.00	36.00	36.00	25.50	25.50	26.00	26.00	22.00	9.00	23.00	35.00	26.00	26.00	26.00		
mPDMS	39.00	18.00	0.00	0.00	21.00	21.00	28.00	28.50	25.50	30.00	28.50		28.00	28.00	28.00		
Norbloc	2.00	2.00	3.00	3.00	2.00	2.00	2.00	2.00	2.00	2.00	2.00	3.00	2.00	2.00	2.00		
CGI 1850	2.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	2.00	1.00	1.00	1.00		
TEGDMA	0.00	0.00	0.00	0.00	1.50	1.50	1.00	1.50	1.50	0.50	1.50			0.25	0.50		
HEMA	0.00	0.00	0.00	0.00	5.00	5.00	5.00	5.00	5.00	7.00	5.00		5.00	5.00	5.00	96.8	98.6
Blue HEMA	0.00	0.00	0.00	0.00	0.02	0.02	0.02	0.02	0.02	0.02	0.02						
PVP	0.00	8.00	0.00	0.00	5.00	5.00	5.00	8.00	5.00	7.50	9.00		5.00				
Darocur 1173																0.3	0.30
EGDMA																0.8	0.8
TMPTMA																0.1	0.1
MAA																2.0	
Diluent %	41	20	20	None	20	50.00	20.00	37.50	20.00	40.00	50.00	20.00	20.00	20.00	20.00	52.00	52.00
Diluent	3M3P	3M3P	3M3P	NA	D30	TAA	D30	3M3P	TAA	3M3P	3M3P	D30	D30	D30	D30	BAGE	BAGE

Example 1

Preparation of Antimicrobial Contact Lenses Using PSPM

5 Contact lenses prepared from PSPM (2365 ppm or 0.24 weight per cent) and base monomer mix (Table 1, 10.0 g). were treated with a 10% w/v solution of AgNO_3 in deionized water for about 60 minutes (30 lenses in 60 mL of 10 wt.% AgNO_3 in deionized water). The treated lenses were removed from the silver solution and placed into distilled water (300 mL). The lenses were
10 either rolled or stirred in distilled water for at least about 20 minutes. This water washing procedure was repeated three (3) more times. The resulting lenses were stored in saline solution and tested to determine their antimicrobial potential. The results of the bacterial adhesion assay are presented in Table 2. In addition, the lenses were analyzed by inductively coupled argon plasma
15 atomic emission spectroscopy (ICP-AES) of a hydrogenfluoride (HF) digest of a dry lens or using instrumental neutron activation analysis, to determine the amount of silver that was incorporated in the lenses. This data is presented in Table 2.

Example 2

Preparation of Antimicrobial Contact Lenses Using Polymerizable Monomers other than PSPM

The procedure of Example 1 was repeated replacing the amount of polymerizable ligand and the base monomer mix as indicated by Table 2.

25 Table 2 lists, the Base Monomer Mix (from Table 1); the polymerizable monomer of Formula I; the concentration of the polymerizable monomer of Formula 1 in ppm; the amount of silver incorporated into the lens; the % inhibition from the bacterial assay, using lenses made from formulation Q without any added polymerizable monomer as the control; the % inhibition from
30 the bacterial assay using lenses made from formulation G as the control. (Lenses were tested by inductively coupled plasma atomic emission spectroscopy). The antimicrobial activity of formulation Q lenses and

formulation G lenses are statistically the same (95% confidence (p=0.05)). In the Base Monomer Mix column, the "Ag" suffix indicates that the lenses were treated with 10% AgNO₃ as described in Example 1.

5

Table 2

no	Base	Monomer of	[Formula I]	[Ag],	% Inhibition	% Inhibition	Trial
	Monomer Mix	Formula I	ppm	ppm	Q	G	
	Q-Ag	PSPM	2365	N/A	96.68	N/A	
10	G	PSPM	2365	**	75.83	52.99	1
	G	PSPM	2365	**	60.64	42.55	2
	G-Ag	PSPM	2365	265	99.64	99.30	1
	G-Ag	PSPM	2365	N/A	97.74	96.71	2
	G	ATU	438	**	20.00	N/A	
15	G-Ag	ATU	438	N/A	45.00	N/A	
	G	ATU	2800	**	0.00	N/A	
	G-Ag	ATU	2800	2700	0.00	N/A	
	G	VIM	1124	**	40.00	N/A	
	G-Ag	VIM	1124	N/A	32.65	N/A	
20	R-Ag	MAA	9,000	15	0.00	0.00	
	R-Ag	MAA	18,000	550	36.85	38.38	
	R-Ag	MAA	36,000	1100	94.89	95.01	
	G	MAA	18,000	**	60.53	46.43	
	G	MAA	27,000	**	36.84	14.29	
25	G	MAA	36,000	**	1.62	0.00	
	G-Ag	MAA	5,000	N/A	46.17	47.48	
	G-Ag	MAA	18,000	N/A	91.34	N/A	
	G-Ag	MAA	27,000	N/A	93.00	N/A	
	G-Ag	MAA	36,000	1800	91.50	N/A	
30	G	MAHB	3610	**	26.32	N/A	
	G	MAHB	16,000	**	27.32	N/A	

		G-Ag	MAHB	3610	1.9	58.70	N/A	
		G-Ag	MAHB	16,000	N/A	13.48	35.15	
		G-Ag	MAHB*	16,000	310	8.36	0.00	
		G	MABP	2512	**	45.22	21.82	
5		G	MABP	89,000	**	0.00	14.34	
		G-Ag	MABP	2512	N/A	53.08	33.03	
		G-Ag	MABP	89,000	1.9	12.13	34.14	
		G-Ag	MABP*	89,000	61	70.03	54.38	
		G	CELL/prot	10,000	**	38.30	58.99	
10		G-Ag	CELL/prot	10,000	3.4	40.73	60.61	
		G	AMPSA	500	**	43.17	0.00	1
		G	AMPSA	1000	**	16.07	0.00	1
		G	AMPSA	1500	**	18.94	0.00	1
		G	AMPSA	2000	**	18.44	0.00	1
15		G	AMPSA	2000	**	29.03	0.00	2
		G	AMPSA	2000	**	10.71	0.00	3
		G	AMPSA	2924	**	5.13	28.50	
		G	AMPSA	3000	**	49.28	22.81	1
		G	AMPSA	3000	**	60.12	30.35	2
20		G	AMPSA	3000	**	13.42	0.00	3
		G	AMPSA	4000	**	0.00	0.00	
		G	AMPSA	5000	**	10.38	0.00	
		G-Ag	AMPSA	500	< 30	67.00	17.69	1
		G-Ag	AMPSA	1000	54.8	59.78	0.00	1
25		G-Ag	AMPSA	1500	296	95.97	89.94	1
		G-Ag	AMPSA	2000	N/A	93.52	90.13	1
		G-Ag	AMPSA	2000	378	98.93	98.13	2
		G-Ag	AMPSA	2000	383	95.02	87.58	3
		G-Ag	AMPSA	2924	1400	97.42	98.02	
30		G-Ag	AMPSA	3000	N/A	93.40	89.96	1
		G-Ag	AMPSA	3000	482	99.13	98.49	2

	G-Ag	AMPSA	3000	150	98.95	98.52	3
	G-Ag	AMPSA	4000	N/A	92.02	87.85	
	G-Ag	AMPSA	5000	N/A	92.25	88.20	
	N	AMPSA	3000	**	65.00	56.20	
5	N-Ag	AMPSA	3000	N/A	98.51	98.14	
	G	CYST	2000	**	64.43	47.39	
	G	CYST	3000	**	61.76	43.45	
	G	CYST	3600	**	0.00	0.00	
	G	CYST	4000	**	55.57	34.30	1
10	G	CYST	4000	**	38.34	0.00	2
	G	CYST	4000	**	8.37	0.00	3
	G	CYST	4000	**	30.90	9.57	4
	G	CYST	5000	**	0.00	0.00	
	G-Ag	CYST	2000	324	90.10	85.35	
15	G-Ag	CYST	3000	436	90.33	85.70	
	G-Ag	CYST	3600	N/A	92.76	94.54	
	G-Ag	CYST	4000	692	93.81	90.84	1
	G-Ag	CYST	4000	725	95.49	88.09	2
	G-Ag	CYST	4000	750	92.64	81.65	3
20	G-Ag	CYST	4000	732	97.22	96.36	4
	G-Ag	CYST	5000	900	85.62	78.73	
	N	CYST	4000	**	36.61	20.67	1
	N	CYST	4000	**	52.82	38.26	2
	N-Ag	CYST	4000	744	94.24	92.79	1
25	N-Ag	CYST	4000	640	99.20	98.96	2
	N-Ag	CYST	4000	719	98.43	97.95	3
	O	CYST	4000	**	67.04	56.87	
	O-Ag	CYST	4000	778	97.00	96.07	
	P	CYST	4000	**	46.51	30.00	
30	P-Ag	CYST	4000	760	98.30	97.77	

N/A indicates data not available

- * indicates that the lenses were prepared by the method of Example 3
- ** indicateds that the lenses were not analyzed for silver content

Example 3

5 Preparation of Antimicrobial Contact Lenses Using Polymerizable Monomers other than PSPM and Treated with a Base Before Silver Treatment

 Contact lenses prepared from a polymerizable monomer (as denoted by an asterix (*) in Table 2; in addition, Table 2 also lists the concentration used) and base monomer mix were treated with either 10% w/v solution of Na_2CO_3 in
10 deionized water, 10% w/v solution of NaHCO_3 in deionized water, 10% w/v solution of NaOH in deionized water, or 1 M NaOMe in methanol for about 10 minutes to about 20 hours (30 lenses in 30 mL of any basic solution). The treated lenses were then removed from the basic solution and placed into deionized water (30 mL). The lenses were either rolled or stirred for a
15 minimum of 10 minutes. This water washing procedure was repeated at least twice. The base treated lenses were then treated with a 10% w/v solution of AgNO_3 in deionized water for about 60 minutes (30 lenses in 60 mL of 10 wt.% AgNO_3 in deionized water). The base/silver treated lenses were removed from the silver solution and placed into distilled water (300 mL). The lenses were
20 either rolled or stirred in deionized water for at least twenty minutes. This water washing procedure was repeated three (3) more times. The resulting lenses were stored in saline solution and tested to determine their antimicrobial potential.

 Table 2 lists, the Base Monomer Mix (from Table 1); the polymerizable
25 monomer of Formula I; the concentration of the polymerizable monomer of Formula 1 in ppm; the amount of silver incorporated into the lens; the % inhibition from the bacterial assay, using lenses made from formulation Q without any added polymerizable monomer as the control; the % inhibition from the bacterial assay using lenses made from formulation G as the control. The
30 antimicrobial activity of formulation Q lenses and formulation G lenses are statistically the same (95% confidence ($p=0.05$)). In the Base Monomer Mix

column, the "Ag" suffix indicates that the lenses were treated with 10% AgNO₃ as described in Example 1.

Example 4

5 Preparation of Antimicrobial Contact Lenses Containing CYST

CYST (0.2 weight percent based on weight of the monomer mix) was polymerized in monomer mix G and cured using the methods outlined in Base Monomer Formulations and Lens Preparation.

10 A mixture of sodium borate (3.70 g) and boric acid (18.52 g) was placed in a 2 L volumetric flask and diluted to volume with deionized water to give Borate Buffered Packing Solution. Silver nitrate (0.1042 g) was weighed into a 100 mL volumetric flask and de-ionized water was added to volume to give Silver Stock Solution. The Silver Stock Solution was further diluted with the Borate Buffered Solution to give a working solution concentration of 0.33 ug
15 Ag/mL. The cured lenses were transferred into vials containing 3 mL of the working solution. The vials were sealed and autoclaved for 2 hours @ 121°C. The treated lenses were removed from the vials and washed with several portions of de-ionized water and subsequently re-packaged in Sodium Chloride Packing Solution (3 mL in vials of 0.85% sodium chloride, 0.9% boric acid,
20 0.18% sodium borate, 0.01% EDTA adjusted to pH of 7.3). The resulting lenses were analyzed for silver content by Instrumental Neutron Activation Analysis (INAA). Lenses produced by this method were characterized by a silver concentration of 46 ug/g.

25

Example 5

Movement of Lenses

Lenses were prepared using the method of Examples 1 2, and 4. The amount of polymerizable ligands and the base monomer are listed. To determine the amount of silver ("[Ag]") present in each lens type, samples were
30 sent Galbraith Laboratories, Inc. (Knoxville, TN) for silver analysis by inductively coupled plasma atomic emission. The first ten lens types in Table 3

were tested on ten (10) subjects per type of lens using the push up assay (Contact Lens Practice, Chapman & Hall, 1994, edited by M. Ruben and M. Guillon, pgs. 589-99). The last six (6) entries in Table 3 were tested on twenty-three (23) subjects per type of lens using the push-up assay. All lenses were evaluated 30 minutes after placing the lenses on patients' eyes. The percentage of lenses having acceptable movement qualities was calculated as follows. Any lens having a score of greater than -2 on the push up test was an acceptable lens. In a each patient study, the number of acceptable lenses was divided by the total number lenses. Lenses having a percentage of movement equal to or greater than 50% are the preferred lenses. In addition, prior to insertions in a patient's eyes the efficacy of the lenses N and G were tested using microbial tests, A and B respectively. The activity of the lenses in these assays is listed in Table 3 as a the log reduction of the assay. Figure 1 shows the percentage lenses having acceptable movement vs the amount of silver in each lens.

Table 3

Base Monomer Mix	Monomer of Formula I	[Formula I] ppm	[Ag] ppm	Log Reduction Assays
N-Ag	CYST	4000	764	1.80
N-Ag ¹	CYST	4000	760	1.60
N-Ag	CYST	4000	823	1.10
N-Ag	CYST	4000	261	1.23
N-Ag	CYST	4000	661	1.48
N-Ag	CYST	4000	212	0.84
N-Ag ²	CYST	4000	790	1.17
G-Ag	CYST	2000	60	1.41
G-Ag	CYST	2000	56	1.45
G-Ag	CYST	2000	164	1.24
G-Ag	CYST	2000	50	2.17
G-Ag	CYST	2000	41	2.24

	G-Ag	CYST	2000	42	2.18
	G-Ag	CYST	2000	49	2.01
	G-Ag	CYST	2000	43	1.91
	G-Ag	CYST	2000	49	1.00
5	G-Ag	APDS	2000	313	1.57

¹ lenses coated with PAA

² lenses coated with pHEMA

10

Example 6

Movement of Lenses

15

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Lenses were prepared using the method of Examples 1 and 2. The amount of polymerizable ligands and the base monomer are listed. To determine the amount of silver (“[Ag]”) present in each lens type, samples were sent Galbraith Laboratories, Inc. (Knoxville, TN) for silver analysis by inductively coupled plasma atomic emission spectroscopy before the lenses were inserted into the eyes of patients. The lens types listed in Table 4 were tested on ten (10) subjects per type of lens using the push up assay (Contact Lens Practice, Chapman & Hall, 1994, edited by M. Ruben and M. Guillon, pgs. 589-99). The lenses were evaluated 30 minutes after placing the lenses on patients’ eyes and the percentage of acceptable movement was calculated as described in Example 5. In addition, the lenses were analyzed for pre-wear antimicrobial efficacy using Test B. The activity of the lenses in these assays is listed in Table 4 as a the log reduction of the assay. Figure 2 shows the percentage lenses having acceptable movement vs the amount of silver in each lens.

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Table 4

Base	Monomer of	[Formula I]	[Ag]	Log Reduction
Monomer Mix	Formula I	ppm	ppm	Assays

Q-Ag	CYST	2000	**	N/A
Q-Ag	CYST	1000	851	N/A
Q-Ag	CYST	500	332	N/A
Q-Ag	CYST	250	58	1.94

5 ** indicates not analyze for silver concentration

Example 7

Movement of Lenses

10 In addition to the testing procedures of Example 5, the lenses of Example 4 were tested after 10 hours, 1 week, 2 weeks and 4 weeks of daily wear use. The percentage of acceptable movement of the lenses was 100%. The lenses were removed from patients' eyes after 10 hours, 1 week, 2 weeks and 4 weeks of use and subsequently analyzed to determine the amount of silver remaining in the lenses. Prior to use, the lenses contained 46 ppm of silver (STD 5). Forty percent (40%) of the silver was lost between 10 hours and one week of daily wear. After one week of daily wear, no further loss of silver was observed.

Example 8

Silver Spike Solution and Lenses

20 CYST (0.4 weight percent based on weight of the monomer mix) was polymerized in monomer mix N and cured using the methods outlined in Base Monomer Formulations and Lens Preparation.

25 Silver nitrate (0.0787 g) was weighed into a 25 mL volumetric flask and Borate Buffered Packing Solution was added to volume to give Solution C ([Ag], 2000 µg/mL). Solution C was further diluted with the Borate Buffered Solution to give the Silver Spike Solution ([Ag], 20 µg/mL). The cured lenses were transferred to vials containing Borate Buffered Packing Solution (3 mL, made by the method of Example 11) and 50µL of Silver Spike Solution was added using an eppendorf pipet. The vials were sealed and autoclaved for 3 consecutive cycles of 30 minutes each @ 121°C. The resulting lenses were
30 analyzed for silver content by Instrumental Neutron Activation Analysis (INAA). The average silver content of the lenses was 45.4 µg/g.

Example 9

Silver Spike Solution and Lenses

5 CYST (0.2 weight percent based on the weight of the monomer mix) was polymerized in monomer mix G and cured using the methods outlined in Base Monomer Formulations and Lens Preparation. A number of lenses were placed in a jacketed beaker, (60 °C) containing 800 mL of sodium borohydride solution (200 µg/mL). The lenses were agitated using a magnetic stirrer for 15 min and subsequently rinsed several times with de-ionized water at 60 °C.

10 Subsequently, the lenses were placed in vials containing Borate Buffered Packing Solution (3 mL, made by the method of Example 11) and 50µL of Silver Spike Solution (Example 8) was added using an eppendorf pipet. The vials were sealed and autoclaved for 3 consecutive cycles of 30 minutes each @ 121°C. The resulting lenses were analyzed for silver content by INAA. The average silver content of the lenses was 44.1 µg/g.

Example 10

Silver Spike Solution and Lenses

20 CYST (02 weight percent based on the weight of the monomer mix) was polymerized in monomer mix G and cured using the methods outlined in Base Monomer Formulations and Lens Preparation. A number of lenses were placed in a jacketed beaker, (60 °C) containing 800 mL of sodium borohydride solution (200 µg/mL). The lenses were agitated using a magnetic stirrer for 15 min and subsequently rinsed several times with de-ionized water at 60 °C.

25 Subsequently, the lenses were placed in vials containing Sodium Chloride Packing Solution (3 mL, of 0.85% sodium chloride, 0.9% boric acid, 0.18% sodium borate, 0.01% EDTA adjusted to pH of 7.3) and 50µL of Silver Spike Solution was added using an eppendorf pipet. The vials were sealed and autoclaved for 3 consecutive cycles of 30 minutes each @ 121°C. The
30 resulting lenses were analyzed for silver content by INAA. The average silver content of the lenses was 44.2 µg/g.

Example 11

Preparation of Antimicrobial Contact Lenses Containing CYST

CYST (0.2 weight percent based on the weight of the monomer mix) was polymerized in monomer mix G and cured using the methods outlined in Base Monomer Formulations and Lens Preparation.

A mixture of sodium borate (1.85 g) and boric acid (9.26 g) was placed in a 1L volumetric flask and diluted to volume with deionized water to give Borate Buffered Packing Solution. Silver nitrate (0.0162 g) was weighed into a 50 mL volumetric flask and de-ionized water was added to volume to give Silver Stock Solution. The Silver Stock Solution was further diluted with the Borate Buffered Solution to give Solution A (1.0 $\mu\text{g/mL}$) and Solution B (0.5 $\mu\text{g/mL}$). The cured lenses were transferred into vials containing 3 mL of either Solution A or Solution B. The vials were sealed and autoclaved for 3 consecutive cycles of 30 minutes each @ 121°C. The resulting lenses were analyzed for silver content by inductively coupled plasma atomic emission spectroscopy. Lenses made using Solution A had an average silver content of 151 $\mu\text{g/g}$. Lenses made using Solution B had an average silver content of 75 $\mu\text{g/g}$.